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Comparison of four seaweed extracts on germination and root promotion of *Lens esculenta*

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ABSTRACT

Biostimulants were considered as biological, organic and synthetic components that promote the plant growth; recently seaweed extracts have been reported as plant biostimulants applied to agricultural systems and there were reported positive effects of them on germination and root promotion. The aim of the present work was to analyze and compare the effect of four seaweed extracts from *Gracilaria debilis*, *Sargassum liebmanni*, *Sargassum vulgare* and *Ulva fasciata* on germination and root elongation of *Lens esculenta*. The plant root elongation promoting (PREP) activity assay was employed to analyze the effect of seaweed liquid extracts on *Lens esculenta* seeds by a tolerance index, the normalized residual percentage of germinated seeds index and the normalized residual elongation root index. Even there was a toxic effect of the extracts from two of the seaweeds tested: *S. vulgare* and *U. fasciata* on *L. esculenta* germination; all the seaweed liquid extracts showed a good response at low concentrations; particularly the employ of *S. liebmannii* and *U. fasciata* are recommended at a moderate dilution rate of their extracts for the increase of germination and growth of plants tested.

Introduction

Biostimulants were considered as biological, organic and synthetic components that promote the plant growth (Sangha et al., 2014); particularly, seaweed mulch or extracts have recently reported as plant biostimulants applied to agricultural systems (Zhang and Schmidt, 1997; Khan et al., 2009; Craigie, 2011; Spann and Little, 2011). Various industrial-scale and commercial processes

have been developed and management as liquid extracts and powders from a wide range of seaweeds such as *Ascophyllum nodosum*, *Fucus* spp., *Laminaria* spp., *Sargassum* spp., *Ecklonia maxima*, and *Durvillaea* spp. (Hong et al., 2007). Some authors have reported that natural seaweeds are used as substitute of synthetic fertilizers; because they contain multiple growth regulators (Begum et al., 2018); such as cytokinins (Durand et al., 2003), auxins (Sahoo, 2000; Stirk et al., 2004),

gibberellins (Wildgoose et al. 1978; Strik and Staden, 1997) and various macro and micronutrients necessary for plant growth and development (Khan et al., 2009). There were some reports regarding to positive effects of seaweed extracts on germination; specially some of them have reported higher germination percentage with low concentration of the extracts. Pramanick et al. (2014) and Layek et al. (2018) reported that when rice seeds were soaked in lower concentrations of seaweed extracts recorded higher germination percentage. Similar results were reported in case of maize plants, when seeds were soaked in 5% concentration of extracts from *Kappaphycus* sp. and *Gracilaria* sp. showing a higher rate of germination; while 15% concentration inhibited the germination (Layek et al., 2016). Higher germination percentage was also reported in *Vigna radiata* (Venkataraman and Mohan, 1997) and *Vigna sinensis* (Sivasankari et al., 2006). Plants treated with seaweed extracts have been reported to show a deep root development by improving lateral root formation (Atzmon and van Staden, 1994) and increased the total volume of the root system (Slàvik, 2005). The aim of the present work was to analyze and compare the effect of four seaweed extracts on germination and root elongation of *Lens esculenta*.

Materials and methods

Preparation of seaweed liquid extracts

For this study, the seaweeds *Sargassum vulgare* and *Ulva fasciata* were hand collected from the intertidal zone at one meter of deep, in “El pulpo beach” located in Barra de Cazonas, from Cazonas de Herrera municipality in Veracruz, México; *Gracilaria debilis* was hand collected from the intertidal zone at one meter of deep, in Villamar, Campeche, México and *Sargassum liebmannii* was were hand collected from the intertidal zone at one meter of deep, in Los Troncos, Guerrero, México. Collected seaweeds were cleaned several times with sea water to remove sand, other impurities and epiphytes, then transported to the laboratory and again cleaned four times with tap water and finally shade dried. These shade-dried seaweeds were finely chopped and powdered with a Nutribullet®. The aqueous extracts were obtained according to Mendoza et al. (2019), as follows: seaweed powders were filtered through

metallic mesh number 16 (1mm) and 25g of each were deposited in flasks filled with 300mL of distilled water and boiled at 80°C for 45 min in a water bath. Then the extracts were allowed to cool at room temperature and filtered through medium pore filter paper. Finally, the filtrates were considered as 100% of seaweed liquid extracts (SLE) and stored at 4°C for the bioassays.

Effect of seaweed extracts on *Lens esculenta* seeds by the plant root elongation promoting (PREP) activity assay

The plant root elongation promoting (PREP) activity assay (Belimov et al., 2005) was employed to analyze the effect of seaweed liquid extracts on *Lens esculenta* seeds. Commercially seeds of *Lens esculenta* were surface-sterilized with 10% sodium hypochlorite solution for 3 minutes and rinsed with deionized sterile water. Fifteen seeds were placed in Petri dishes with sterile filter paper; 7mL of the different concentrations of each SLE: 5, 10, 20 and 30% prepared by diluting the concentrate extracts with sterile distilled water, were added and 7mL of sterile distilled water was considered as control. All the experiments were performed by triplicate and maintained in a growth chamber incubated in dark at 28°C for 5 days. After this period, seeds germinated were counting and seedlings were obtained and photographed. The root length of each plant was measured employing the EZ-Rhizo II (Ver. 2.4.5.10) program.

The analysis of the effect of seaweed extracts were done measuring the germination percentage and root length of *L. esculenta* seedlings after their incubation; tolerance index (TI) was obtained as the ratio of the root lengths of seedlings grown in the presence and absence of the specific added seaweed extract (Wilkins, 1978; Burd et al., 1998); $TI = RL_{se} / RL_c$ where RL_{se} is the root length of plants grown in the presence of a specific added seaweed extract and RL_c is the root length of plants grown in absence of seaweed extract (control). Also the toxicity of experimental conditions to *L. esculenta*, was measured according to Bagur-González et al. (2011), by the Normalized Residual Percentage of Germinated Seeds Index (NRPGI) following the equation: $NRPGI = G_{exp} - G_c / G_c$ Where, G_{exp} is the % average of germinated seeds in experimental conditions tested (each seaweed extracts at 5, 10, 20 and 30%), and G_c is the %

average of germinated seeds in control condition and by the Normalized Residual Elongation Root Index (NRERI) following the equation: $NRERI = RE_{exp} - RE_c / RE_c$ Where, RE_{exp} is the average length of the root seedlings in experimental conditions tested and RE_c is the average length of the root seedlings in control condition. Bagur-González et al. (2011) noted that the employ of these indices allows the analysis in this study of the toxicity produced by the SLE tested; with values between -1 (maximum phytotoxicity) to > 0 , in a scale proposed by these authors that gives a good classification of this effect: A values = 0 to -0.25 (low toxicity), B values = -0.25 to -0.5 (moderate toxicity), C values = -0.5 to -0.75 (high toxicity) and D values = -0.75 to -1 (very high toxicity); values > 0 indicated a stimulation of growth determined as hormesis effect.

Statistical analysis

All data obtained were analyzed by one-way analysis of variance and the mean differences were compared applying a Tukey-Kramer Method using the statistics program Graph Pad InStat Ver. 2.03. A numerical comparative analysis considering the experimental conditions; a distance matrix was built using the conventional standard distance coefficient, a phenogram was built using the unweighted pair group method of arithmetic averages (UPGMA) method and correlation coefficient of Pearson was obtained using the NTSyS-PC version 2.11T (Numerical Taxonomy and Multivariate Analysis System) software.

Results and discussion

Effect of seaweed extracts on *Lens esculenta* germination and root promotion

Fig. 1 shows the *L. esculenta* seedlings comparing all the seaweed extracts tested in each Petri dish bioassays. Fig. 2 shows the particularly details about the roots growth in each experimental conditions. Germination percentage of *L. esculenta* seeds, showed a particularly positive effect of *S. liebmannii* and *G. debilis* with 100%; while this percentage reduced in seeds exposed to *U. fasciata* as follows: $100 > 83 > 17 > 16\%$ with 5, 10, 20 and 30% extract concentrations and seeds exposed to *S. vulgare* liquid extracts: $51 > 49 > 44 > 40$ with 20, 5, 10 and 30% concentrations. Fig. 3 presented

the root length obtained in all the experimental SLE concentrations tested; according to the obtained results, the comparison between the root lengths in each particular condition showed that even all the seaweed extracts promote the root elongation below the control response of roots; the best elongation response was obtained in the next order: *U. fasciata* $>$ *S. liebmannii* $>$ *S. vulgare* $>$ *G. debilis*.

Phytotoxicity of seaweed extracts tested regarding to germination and root response of *L. esculenta*

The response against the seaweeds extracts, was determined by the RTI (Table 1), against SLE where experiments with *S. liebmannii* extracts showed the highest values of roots tolerance: 0.73, 0.97, 0.67 and 0.85 to 5, 10, 20 and 30%, respectively; followed by *U. fasciata* extract with TI values: 0.67, 0.73 and 0.58 for 5, 10 and 20%, respectively. Almost the same root inhibitory response was obtained between all the concentrations tested of liquid extracts from *S. vulgare* and *G. debilis*, where no particular differences between them were obtained (0.33 to 0.50). As Corona et al. (2018) reported the results of these experiments presented as tolerance index are important according also to Burd et al. (1998); where a TI value of 1.0 indicates that the experiment condition was not inhibitory, whereas a value of 0.1 indicates this was only 10% of the growth of control seedlings.

Regarding to the possible toxic effect of seaweeds extracts on *L. esculenta*, the analysis done of the phytotoxic effect by the determination of the NRPGI and NRERI indices according to Bagur-González et al. (2011) (Table 2), showed that experiments that affect seed germination were grouped in four categories: "A" (low toxicity): *G. debilis* and *S. liebmannii* at all the concentrations tested, *U. fasciata* at 5 and 10% and *S. vulgare* at 20%; "B" (moderate toxicity): *S. vulgare* at 5%; "C category" (high toxicity): *S. vulgare* at 10 and 30%; and "D" (very high toxicity): *U. fasciata* at 20 and 30%. Regarding to the effect on seed germination, no toxicity at all was founded and the SLE concentrations tested for *S. liebmannii* and *G. debilis* promotes the 100% of germination. For *S. vulgare* and *U. fasciata* SLE concentrations of 20 and 10% respectively promotes the germination of

L. esculenta agree with the results reported by Kumar and Sahoo (2011), whose noted that the application of *Sargassum wightii* seaweed liquid extract increases the seed germination percentage of *Triticum aestivum* var. Pusa Gold. Seeds treated with 20% seaweed extract gave highest percentage of germination. Xavier and Jesudass (2007) also reported that 100% seed germination was found in lower concentrations of *Caulerpa recemosa* extract. Jothinayagi and Anbazhagan (2009) also reported the effect of different concentrations of SLE of *Sargassum wightii* on germination percentage of *Abelmoschus esculentus*; where seeds germination (100%) was found at 20% concentration. *A. esculentus* seeds soaked with

lower concentrations of the *S. wightii* extracts showed higher rates of germination, while the higher concentrations of the extracts inhibited the germination. Sivasankari et al. (2006) analyzed the effect of liquid extracts from *Sargassum wightii* and *Caulerpa chemnitzia* on germination percentage of *Vigna sinensis*; for both seaweeds, greatest seed germination (100 and 98%, respectively) was found at 20% concentration in SLE soaked seeds. Challen and Hemingway (1965) noted that the increase of germination percentage at low concentrations may be due to the presence of some growth promoting substances (such as auxins, gibberellins and cytokinins), micronutrients, vitamins and amino acids.



Fig. 1: *Lens esculenta* seedlings exposed to the four seaweed extracts: **G:** *Gracilaria debilis*, **Sli:** *Sargassum liebmannii*, **Sv:** *Sargassum vulgare* and **U:** *Ulva fasciata*.



Fig. 2: *Lens esculenta* root development under the four seaweed extracts: **G:** *Gracilaria debilis*, **Sli:** *Sargassum liebmannii*, **Sv:** *Sargassum vulgare* and **U:** *Ulva fasciata*.

The germination percentage increased with concentration levels upto 20% and there after it declined. No germination was found at 50% and above. The lowest germination percentage (65%) was found at 40% extract of *C. chemnitzia* treated water soaked seeds. Similar results were obtained for *L. esculenta* treated with *U. fasciata* 30% liquid extract, classified as a very toxic seaweed extract.

Experiments that influenced root length were characterized according to “A category” (low toxicity): *S. liebmannii* only at 10 and 30%; “B

category” (moderate toxicity): *S. liebmannii* only at 5 and 20%, *U. fasciata* at 5, 10 and 20% and *S. vulgare* at 20%; and “C category” (high toxicity): *G. debilis* at all the concentrations tested, *S. vulgare* at 5, 10 and 30% and *U. fasciata* at 30%. Regarding to root elongation, *Sargassum* species tested showed that all *S. liebmannii* SLE concentrations tested and particularly the SLE *S. vulgare* 20% concentration, promoted the root response. Even *U. fasciata* showed moderate toxicity for all its LSE concentrations tested; there was a promotion of root length at 5, 10 and 20%.

Table 1. Determination of Tolerance Index of *Lens esculenta*.

SLE (%)	Root Tolerance Index
<i>Sargassum liebmannii</i>	
5	0.737
10	0.977
20	0.676
30	0.858
<i>Sargassum vulgare</i>	
5	0.471
10	0.484
20	0.501
30	0.43
<i>Gracilaria debilis</i>	
5	0.417
10	0.326
20	0.479
30	0.339
<i>Ulva fasciata</i>	
5	0.676
10	0.732
20	0.589
30	0.351

Relationship between experimental conditions and *L. esculenta* response

Finally, according to the multivariate analysis that consider germination percentage, root length and root tolerance index as parameters; Fig.4 shows the association of groups according with the nature of the responses obtained for each experimental condition; where two groups forming at first: group Ia made only by the 10% LSE from *U. fasciata* and group Ib by all the SLE concentrations from *S. liebmannii* and *G. debilis* and SLE of 5% from *U. fasciata*; the rest of them comprise the group II; in this group the control *L. esculenta* response and LSE concentrations 20 and 30% from *U. fasciata* form the group IIa and the other conditions forming the group IIb by all SLE concentrations from *S. vulgare*.

This analysis demonstrate that there were some particularly relationships between *S. vulgare* and *U. fasciata* from SLE of 20 and 30% concentrations that were similar and the other seaweeds tested also showed a particular association between *G. debilis* and *S. liebmannii* as more closer groups.

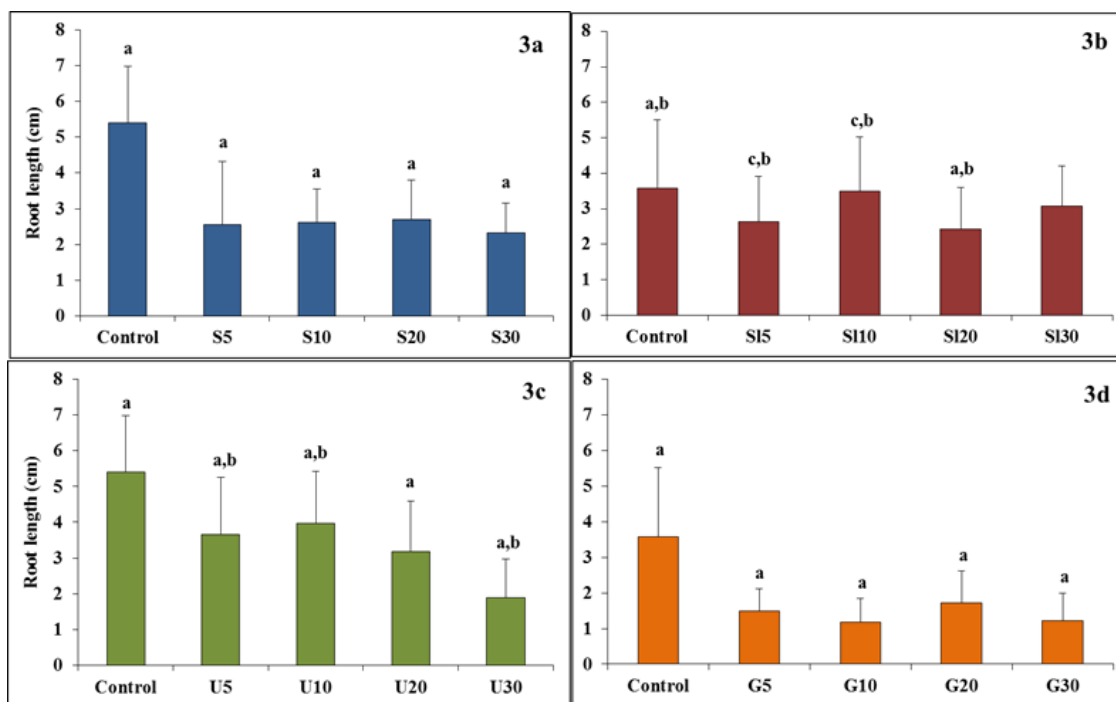


Fig.3. *Lens esculenta* root length: **3a)** *Sargassum vulgare* liquid extract, **3b)** *Sargassum liebmannii* liquid extract, **3c)** *Ulva fasciata* liquid extract and **3d)** *Gracilaria debilis* liquid extract. [n = 15. Mean values \pm S.D. from triplicates are given. The different lower-case letters shows the significant differences founded (a= $p < 0.001$, b= $p < 0.01$, c= $p < 0.05$)].

Table 2. Determination of NRERI and NRPGI indices of *Lens esculenta* seedlings.

NRERI index				
SLE (%)	<i>Sargassum liebannii</i>	<i>Sargassum vulgare</i>	<i>Gracilaria debilis</i>	<i>Ulva fasciata</i>
5	B, -0.2625	C, -0.5284	C, -0.5821	B, -0.3235
10	A, -0.0223	C, -0.5157	C, -0.6735	B, -0.2673
20	B, -0.3234	B, -0.4984	C, -0.5208	B, -0.4106
30	A, -0.1418	C, -0.5697	C, -0.6609	C, -0.6488
NRPGI index				
SLE (%)	<i>Sargassum liebannii</i>	<i>Sargassum vulgare</i>	<i>Gracilaria debilis</i>	<i>Ulva fasciata</i>
5	A, 0	B, -0.50005	A, 0	A, 0
10	A, 0	C, -0.5556	A, 0	A, -0.1668
20	A, 0	A, -0.1867	A, 0	D, -0.8223
30	A, 0	C, -0.5927	A, 0	D, -0.8397

*Where: A= 0 a -0.25 low toxicity, B= -0.25 a -0.5 moderate toxicity, C= -0.5 a -0.75 high toxicity, D = -0.75 a -1.0 very high toxicity y E = > 0 hormesis (Bagur-González et al., 2011).

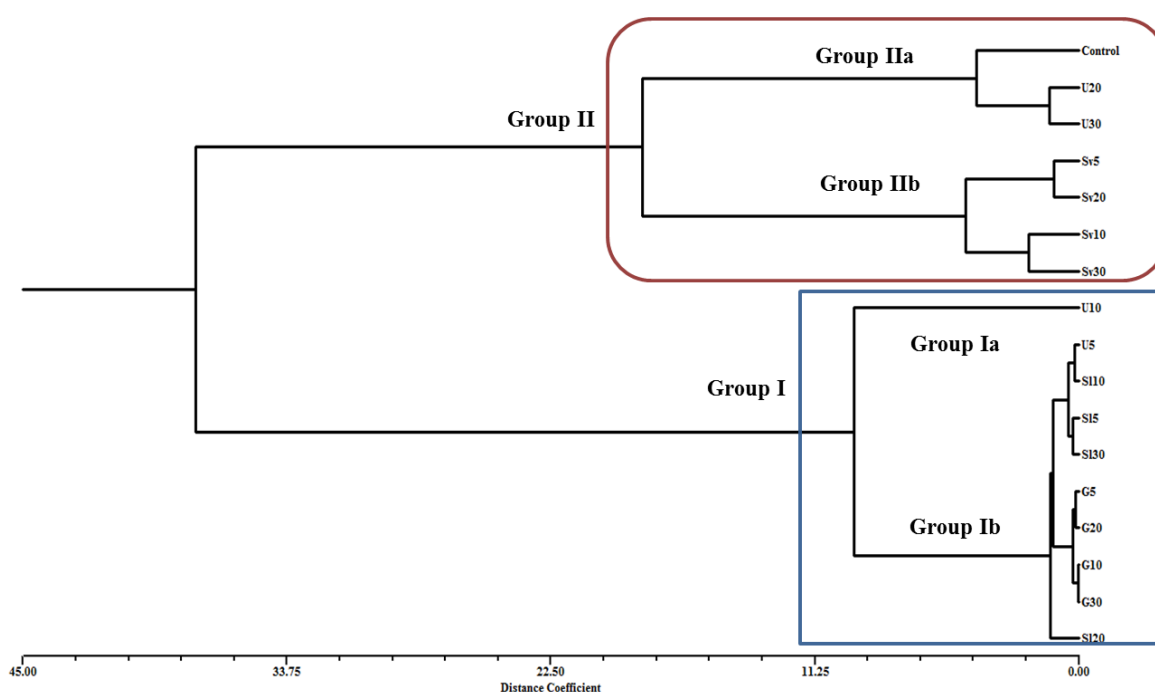


Fig. 4: Phenogram comparing the experimental conditions tested (r= 0.91).

Conclusion

This work showed the particularly effect of four seaweed extracts tested from *Gracilaria debilis*, *Sargassum liebmanni*, *Sargassum vulgare* and *Ulva fasciata* on germination and root length response of *Lens esculenta*, where even there was a toxic effect of the extracts from two of the seaweeds tested: *U. fasciata* and *S. vulgare* on *L. esculenta* germination; all the seaweed liquid extracts showed a good response at low concentrations; particularly the employ of *S. liebmannii* and *U. fasciata* are recommended at

moderate dilution rate of their extracts for the increase of germination and growth of plants tested.

Conflict of interest statement

Authors declare that they have no conflict of interest.

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References

- Atzmon, N., van Staden, J., 1994. The effect of seaweed concentrate on the growth of *Pinus pinea* seedlings. *New For.* 8, 279-288.B
- Bagur-González, M.G., Estepa-Molina, C., Martín-Peinado, F., Morales-Ruano, S., 2011. Toxicity assessment using *Lactuca sativa* L. bioassay of the metal (loid)s As, Cu, Mn, Pb and Zn in soluble-in-water saturated soil extracts from an abandoned mining site. *J. Soils Sediments.* 11, 281-289.
- Begum, M. Bordoloi, B.Ch., Singha, D.D., Ojha, N.J., 2018. Role of seaweed extract on growth, yield and quality of some agricultural crops: a review. *Agricult. Rev.* 39, 321-326.
- Belimov, A.A., Hontzeas, N., Safronova, V.I., Demchinskaya, S.V., Piluzza, G., Bullitta, S., Glick, B.R., 2005. Cadmium-tolerant plant growth-promoting bacteria associated with the roots of Indian mustard (*Brassica juncea* L. Czern.). *Soil Biol. Biochem.* 37, 241-250.
- Burd, G.I., Dixon, D.G., Glick, B.R., 1998. A plant growth-promoting bacterium that decreases nickel toxicity in seedlings. *Appl. Environ. Microbiol.* 64, 3663-3668.
- Challen, S.B., Hemingway, J.C., 1965. Growth of higher plants in response to feeding with seaweed extracts. *Proc. Int. Seaweed Symp.* 5, 359-367.
- Corona-Álvarez, D., Guerrero-Zúñiga, L.A., Rodríguez-Dorantes, A., 2018. Comparison in growth parameters of the bacterization effect on *Axonopus affinis* seeds and EDTA against cadmium. *Int. J. Curr. Res. Biosci. Plant Biol.* 5, 25-34.
- Craigie, J.S., 2011. Seaweed extracts stimuli in plant science and agriculture. *J. Appl. Phycol.* 23, 371-393.
- Durand, N., Brian, X., Meyer, C., 2003. The effect of marine bioactive substances (N PRO) and endogenous cytokinins on nitrate reductase activity in *Arabidopsis thaliana*. *Physiol. Plant.* 119, 489-493.
- Hong, D., Hien, H., Son, P., 2007. Seaweeds from Vietnam used for functional food, medicine and biofertilizer. *J. Appl. Phycol.* 19, 817-826.
- Jothinayagi, N., Anbazhagan, C., 2009. Effect of seaweed liquid fertilizer of *Sargassum wightii* on the growth and biochemical characteristics of *Abelmoschus esculentus* (L.) Medikus. *Recent Res. Sci. Technol.* 1, 155-158.
- Khan, W., Rayirath, U.P., Subramanian, S., Jithesh, M.N., Rayorath, P., Hodges, D.M., Critchley, A.T., Craigie, J.S., Norrie, J., Prithiviraj, B., 2009. Seaweed extracts as biostimulants of plant growth and development. *Plant Growth Regul.* 28, 386-399.
- Kumar, G., Sahoo, D., 2011. Effect of seaweed liquid extract on growth and yield of *Triticum aestivum* var. Pusa Gold. *J. Appl. Phycol.* 23, 251-255.
- Layek, J., Das, A., Ramkushna, G.I., Ghosh, A., Panwar, A.S., Krishnappa, R., Ngachan, N.V., 2016. Effect of seaweed sap on germination, growth and productivity of maize (*Zea mays*) in North Eastern Himalayas. *Indian J. Agron.* 61, 354-359.
- Layek, J., Das, A., Ramkrushna, G.I., Sarkar, D., Ghosh, A., Zodape, S.T., Lal, R., Yadav, G.S., Panwar, A.S., Ngachan, S., Meena, R.S., 2018. Seaweed extract as organic bio-stimulant improves productivity and quality of rice in eastern Himalayas. *J. Appl. Phycol.* 30, 547-558.
- Layek, J., Ramkrushna, G.I., Das, A., Ghosh, A., Krishnappa, R., Panwar, A.S., Azad Thakur, N.S., Ngachan, S.V., Zodape, S.T., Buragohain, J., Mawlong, B., 2014. Seaweed sap as organic iostimulant for rice and maize production. *Research Bulletin No.82. ICAR Research Complex for NEH region, Umiam, Meghalaya, India.*
- Mendoza-Morales, L.T., Mendoza-González, A.C., Mateo- Cid, L.E., Rodríguez-Dorantes, A., 2019. Effect of seaweed liquid extracts on the internode variation of *Lens esculenta* seedlings. *Int. J. Sci.* 8, 1-5.
- Pramanick, B., Brahmachari, K., Ghosh, A., 2014. Efficacy of *Kappaphycus* and *Gracilaria* sap on growth and yield improvement of sesame in new alluvial soil. *J. Crop Weed.* 10, 77-81.
- Sahoo, D., 2000. Farming the ocean. In: *Seaweeds Cultivation and Utilization*. Aravali Books International. New Delhi, India, 40.

- Sangha, J.S., Kelloway, S., Critchley, A.T., Prithiviraj, B., 2014. Seaweeds (Macroalgae) and their extracts as contributors of plant productivity and quality: the current status of our understanding. *Adv. Bot. Res.* 71, 189-219.
- Sivasankari, S., Venkatesalu, V., Anantharaj, M., Chandrasekaran, M., 2006. Effect of seaweed extracts on the growth and biochemical constituents of *Vigna sinensis*. *Bioresour. Technol.* 97, 1745-1751.
- Slàvik, M., 2005. Production of Norway spruce (*Picea abies*) seedlings on substrate mixes using growth stimulants. *J. For. Sci.* 51, 15-23.
- Spann, T.M., Little, H.A., 2011. Applications of a commercial extract of the brown seaweed *Ascophyllum nodosum* increases drought tolerance in container-grown 'Hamlin' sweet orange nursery trees. *HortSci.* 46, 577-582.
- Stirk, W.A., Arthur, G.D., Lourens, A.F., Novak, O., Strnad, M., van Staden, J., 2004. Changes in cytokinin and auxin concentrations in seaweed concentrates when stored at an elevated temperature. *J. Appl. Phycol.* 16, 31-39.
- Strik, W., Staden, V.J., 1997. Isolation and identification of cytokinins in a new commercial seaweed product made from *Fucus serratus* L. *J. App. Phycol.* 9, 327-330.
- Venkataraman, K.V., Mohan, V.R., 1997. Effect of seaweed extract SM3 on the cyanobacterium, *Scytonema* species. *Seaweed Res. Utiln.* 19, 13-15.
- Wildgoose, P.B., Blunden, G., Jewers, K., 1978. Seasonal variation in gibberellin activity of some species of Fucaceae and Laminariaceae. *Bot. Mar.* 21, 63-65.
- Wilkins, D.A., 1978. The measurement of tolerance to edaphic factors by means of root growth. *New. Phytol.* 80, 623-633.
- Xavier, G.S.A., Jesudass, L.L., 2007. Effect of seaweed extracts on cluster bean. *Seaweed Res. Util.* 29, 85-87.
- Zhang, X., Schmidt, R., 1997. The impact of growth regulators on the α -tocopherol status in water-stressed *Poa pratensis*. *Int. Turfgrass Soc. Res. J.* 8, 1364-1371.

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